

Protecting and improving the nation's health

Investigation of novel SARS-CoV-2 variant

Variant of Concern 202012/01

Technical briefing 2

This briefing provides an update on the <u>briefing</u> of 21/12/2020.

Nomenclature of variants in the UK

SARS-CoV-2 variants if considered to have concerning epidemiological, immunological or pathogenic properties are raised for formal investigation. At this point they are designated Variant Under Investigation (VUI) with a year, month, and number. Following risk assessment with the relevant expert committee, they may be designated Variant of Concern (VOC). This variant was designated VUI 202012/01 on detection and on review re-designated as VOC 202012/01 on 18/12/2020.

Current epidemiological findings

Only a small fraction of all new cases of VOC 202012/01 are identified by wholegenome sequencing, and this data typically lags test date by approximately two weeks, therefore a proxy S gene target failure (SGTF)) is used to indicate carriage of the VOC.

We previously observed that one of the S gene mutations in the VOC, which deletes amino acids 69 and 70 (Δ 69-70), causes a reproducible SGTF in the Thermopath TaqPath assay used in three UK lighthouse laboratories (see <u>Technical Briefing 1</u>). This coincidental occurrence therefore provides a good proxy for monitoring trends in VOC 202012/01. SGTF correlates almost perfectly with presence of Δ 69-70. Considering 14,950 tested samples where we know both the sequence and the SGTF status, 99.3% of Δ 69-70 sequences (1,831 of 1,843) are SGTF, compared to 0.05% of sequences without the deletion (7 of 13107).

Because $\Delta 69$ -70 has arisen multiple times, and SGTF is a proxy for any lineage with that mutation, the utility of SGTF as a proxy for VOC 202012/01 varies over time and region. Table 2 shows, for all pillar 2 sequences, the weekly proportion of $\Delta 69$ -70 sequences that were confirmed to be VOC 202012/01. Table 3 shows the proportion of $\Delta 69$ -70 that is the VOC 202012/01 in England during December, broken down by region. It is, as expected, highest in the areas where the VOC was first observed, but it has been a substantial majority in all areas of England during the month of December. The numbers in these tables are based on sequenced samples, some of which may have come from the same individual (this effect is likely to be small).

Table 1. Proportion of all Pillar 2 $\Delta 69-70$ sequences that are VOC-202012/01 by week.

Week beginning	Per cent VOC of all Δ69-70	Number of pillar 2 Δ69-70 sequences
2020-10-12	3%	116
2020-10-19	15%	219
2020-10-26	29%	156
2020-11-02	64%	398
2020-11-09	79%	632
2020-11-16	88%	602
2020-11-23	93%	372
2020-11-30	96%	370
2020-12-07	98%	2,007

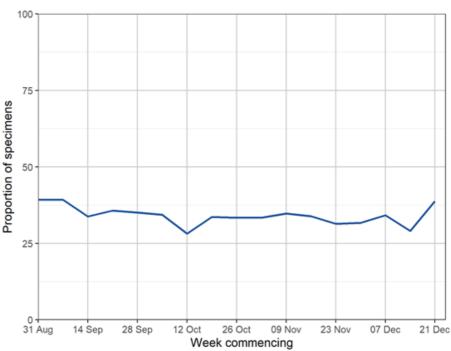
Table 2. Proportion of all Pillar 2 $\Delta 69\text{--}70$ sequences that are VOC 202012/01 by region.

Region	Per cent VOC of all Δ69-70	Number of pillar 2 Δ69-70 sequences December 1 st -21 st
East Midlands	82%	45
East of England	99%	375
London	98%	1056
North East	91%	58
North West	93%	83
South East	99%	646
South West	100%	34
West Midlands	98%	80
Yorkshire and the Humber	89%	36

The proportion of England specimens tested in the three lighthouse laboratories using the assay which produces the S gene drop out is substantial and relatively constant over time (Figure 1). The proportion of cases tested by this assay which are SGTF has continued to rise in December, in all age groups though most markedly in persons aged 25-49 years. The spatial distribution of SGTF cases shows a relatively higher burden in the South East and London, parts of the South West region and Cumbria.

To note, the low coverage in East of England by the three Thermofisher TaqPath labs precludes any analysis of SGTF burden in that region. There is less confirmatory genomic data in the North West and additional sequencing is being undertaken

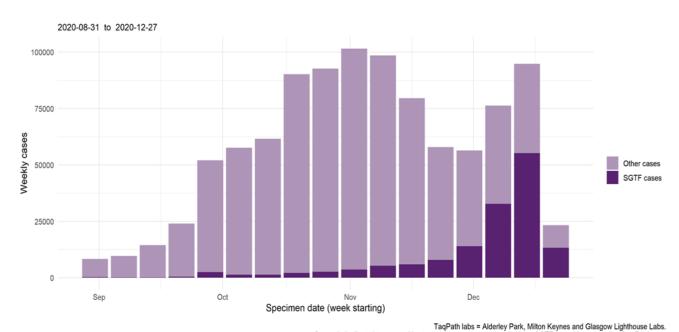
Figure 1. Proportion of England specimens tested in Thermofisher TaqPath, by S-gene detection by week, 1 September to 22 December 2020



TaqPath Labs = Alderley Park, Milton Keynes and Glasgow Lighthouse Labs Includes both positive and negative SARS-CoV specimens from Pillar 1 and 2.

Excludes lateral flow device tests. Data source: USD

Figure 2. Weekly number of cases tested by laboratories using Thermofisher TaqPath, by S-gene detection

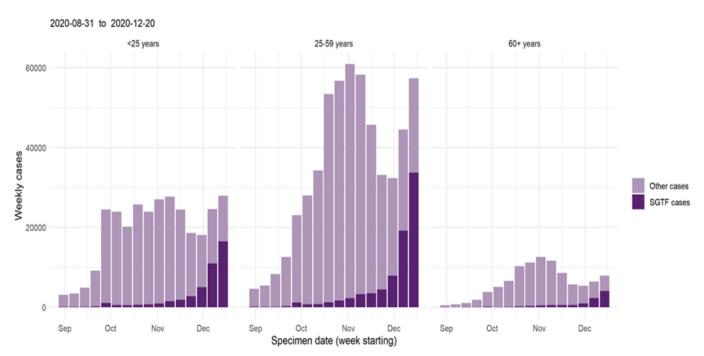


TaqPath labs = Alderley Park, Milton Keynes and Glasgow Lighthouse Labs.

Cases deduplicated to one positive test per person per week, prioritising SGTF tests. Natural weeks Mon-Fri shown.

Data source: SGSS

Figure 3. Weekly number of cases tested by laboratories using Thermofisher TaqPath, by S-gene detection and age group

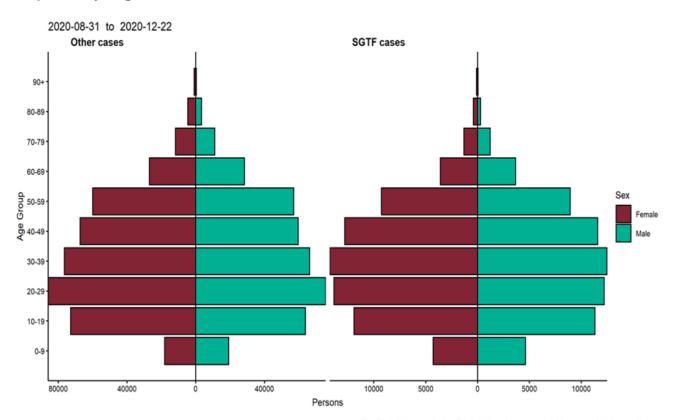


TaqPath labs = Alderley Park, Milton Keynes and Glasgow Lighthouse Labs.

Cases deduplicated to one positive test per person per week, prioritising SGTF tests. Natural weeks Mon-Fri shown.

Data source: SGSS. Age missing in 5257 persons, excluded from figure.

Figure 4. Age-sex pyramid of cases tested by laboratories using Thermofisher TaqPath, by S-gene detection

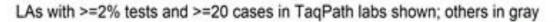


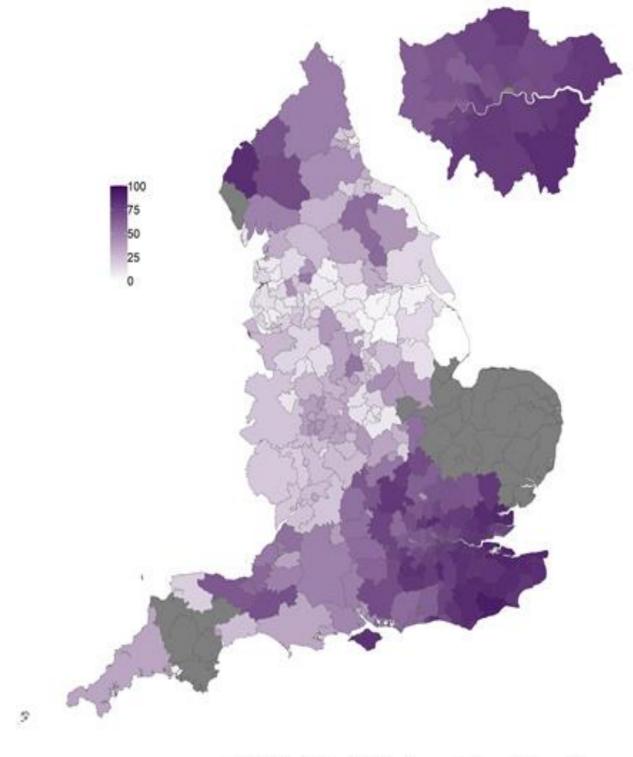
TaqPath labs = Alderley Park, Milton Keynes and Glasgow Lighthouse Labs.

Cases deduplicated to one positive test for entire time period, prioritising SGTF tests where individuals test positive multiple times.

Data source: SGSS. Age and sex missing in 5227 persons, excluded from figure.

Figure 5. Proportion of COVID-19 cases with SGTF among cases tested by laboratories using Thermofisher TaqPath, by S-gene detection (14 to 20 December 2020)





TaqPath labs = Alderley Park, Milton Keynes and Glasgow Lighthouse Labs.

Cases deduplicated to one positive test per person per week, prioritising SGTF tests. Natural weeks Mon-Sun shown.

Data source: SGSS

Preliminary findings of matched cohort study

A matched cohort study was undertaken to inform a preliminary assessment of outcomes of hospitalisation and case fatality associated with VOC 202012/01. This analysis of sequenced positive SARS-CoV2 cases used confirmed VOC cases matched to confirmed wild-type comparator cases (classified as distinct to the variant sequence). To optimise comparability of the variant (VOC 202012/01) and wild-type cases and manage the impact of non-random sampling of SARS-CoV-2 cases for sequencing, variant cases were frequency matched to wild-type cases on a 1:1 basis by age group, sex, upper tier local authority (UTLA) of residence and two-week time-period for specimen date.

Of the 2,693 variant cases identified at the time of analysis, 1,769 variant cases with specimen dates between 20 September and 15 December 2020 were matched to 1,769 wild-type comparator cases and were included in this analysis. In reflection of the matching criteria, the median age of variant cases was 36 years and 35 for the wild-type cases. 51.4% of both the variant cases wild-type comparator cases were female.

The comparison of the 3,538 variant and wild-type cases showed that the majority of variant cases were of White ethnicity (75.2%) followed by Asian (10.1%) and Black (5.9%). The ethnic profile of wild-type comparator cases was broadly similar but a higher proportion of Asian ethnicity (13.5% v 10.1%).

The majority of variant cases were resident in private dwellings (95.0% in variant cases and 94.3% in wild-type comparator cases). Variant cases were more likely to be part of a residential cluster (defined as all laboratory confirmed cases occurring at the same Unique Property Reference Number (UPRN) within 14 days of each other) compared to wild-type comparator cases (63.5% vs 56.1%, Chi-Squared test p=0.00).

Review of hospital admissions data from the NHS identified that of the 3,538 cases, 42 individuals had a record of hospital admission after the date of specimen. Fewer variant cases (16 cases (0.9%)) were admitted to hospital compared to wild-type comparator cases (26 cases (1.5%)) but the difference was not significant (Chi-squared test p=0.162). Due to potential time delays for receipt of hospital admissions data, the identified hospital admissions should be regarded as a minimum number of hospital admissions and further admissions data are likely to be received into this NHS dataset in the future.

The 28-day case fatality was assessed for variant cases and comparator cases. Analysis was restricted to 2,700 cases with a full 28 days elapsed since the specimen date. Among variant cases, 12 of 1,340 (0.89%) variant cases died within 28 days of their specimen date compared with 10 of 1,360 (0.73%) wild-type comparator cases; this difference was not signflicant (Odds ratio:1.21, p=0.65).

Re-infection

Laboratory data were used to identify possible reinfections; these were defined as an episode of polymerase chain reaction (PCR) positivity at least 90 days before a recent PCR positive detection. Two reinfections were detected in in the variant case group (1.13/1000 cases) compared to 3 reinfections in the comparator group (1.70/1000 cases, Fisher's exact P=1.00).

The same definition was applied to SGTF cases. These SGTF cases included samples with orf and N gene Ct values of <31, S gene defined as negative or positive based on presence or absence regardless of Ct value. The rate of detected re-infections in national SGTF cases was 0.60/1000 compared to a rate of 0.61/1000 in non-SGTF cases (P=0.94). When limited geographically to the Kent area, the rate of detected re-infections was 0.51/1000 compared to 0/1000 in non-SGTF cases (P=0.69).

Secondary attack rates

We investigated secondary attack rates using data from NHS Test and Trace, the national contact tracing system in England. Between 5 October and 6 December 2020, 1,105,388 cases were reported to NHS Test and Trace; 46,237 (4.2%) had genomic sequencing data (around 700 (1-2%) poor quality). 1,978 had the variant (VOC 202012/01), 4.3% of those with sequencing data.

228,361 (9.9% attack rate) of all contacts notified by cases in this period became cases:

- 15.1% among those whose index case was confirmed to have the VOC 202012/01
- 9.8% among those whose index case was sequenced and confirmed with other variants

Summary

The findings from the analysis of data on SGTF cases showed a roughly similar spatial distribution of cases as observed from mapping of genomic data. SGTF cases were mostly observed in the South East, London parts of the South West regions and Cumbria. This regional variation in should be interpreted in the context of limited coverage in regions like East of England of by the three Thermofisher TaqPath lighthouse labs. However, in regions with relatively high and consistent coverage, the findings can be used as a proxy for the burden of VOC 201212/01 infection.

Preliminary results from the cohort study found no statistically significant difference in hospitalisation and 28-day case fatality between cases with the variant (VOC 201212/01) and wild-type comparator cases. There was also no significant difference in the likelihood of reinfection between variant cases and the comparator group.

Data sources

Data used in this investigation is derived from the COG-UK dataset, the PHE Second Generation Surveillance System, the secondary uses service (SUS) dataset and Emergency Care Data Set (ECDS).

GISAID reference genome

Sequences from this VOC can be identified by searching for the B1.1.7 lineage on GISAID (gisaid.org). The canonical VOC genome is deposited with accession EPI ISL 601443.

Contact: All enquiries relating to scientific or public health matters should be addressed to PHE.NICC34@phe.gov.uk

About Public Health England

Public Health England exists to protect and improve the nation's health and wellbeing, and reduce health inequalities. We do this through world-leading science, research, knowledge and intelligence, advocacy, partnerships and the delivery of specialist public health services. We are an executive agency of the Department of Health and Social Care, and a distinct delivery organisation with operational autonomy. We provide government, local government, the NHS, Parliament, industry and the public with evidence-based professional, scientific and delivery expertise and support.

Public Health England Wellington House 133-155 Waterloo Road London SE1 8UG Tel: 020 7654 8000

www.gov.uk/phe
Twitter: @PHE_uk

Facebook: www.facebook.com/PublicHealthEngland

Authors

PHE: Meera Chand, Susan Hopkins, Gavin Dabrera, Hester Allen, Theresa

Lamagni, Obaghe Edeghere, Christina Achison, Richard Myers

Imperial College London: Wendy Barclay, Neil Ferguson, Erik Volz

University of Birmingham: Nick Loman University of Edinburgh: Andrew Rambaut Wellcome Sanger Institute: Jeff Barrett

Acknowledgements

The authors are grateful to those teams and groups providing data for this analysis, including: the Lighthouse Laboratories, COG-UK, the Wellcome Sanger Institute, the PHE Epidemiology Cell and outbreak surveillance team.

For queries relating to this document, please contact: PHE.NICC34@phe.gov.uk



© Crown copyright 2020

You may re-use this information (excluding logos) free of charge in any format or medium, under the terms of the Open Government Licence v3.0. To view this licence, visit <u>OGL</u>. Where we have identified any third party copyright information you will need to obtain permission from the copyright holders concerned.

Published December 2020 PHE gateway number: GOV-7132



PHE supports the UN Sustainable Development Goals

